

CERTIFICATE OF ANALYSIS 384043

Client Details

Client	NSW Health
Attention	Kwendy Cavanagh
Address	Locked Bag 2030, ST LEONARDS, NSW, 1590

Sample Details

Your Reference	<u>Narrabri Shire - Namoi Reservoir N27</u>
Number of Samples	3 Water
Date samples received	24/06/2025
Date completed instructions received	24/06/2025

Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.
Samples were analysed as received from the client. Results relate specifically to the samples as received.
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.
Please refer to the last page of this report for any comments relating to the results.

Report Details

Date results requested by	25/06/2025
Date of Issue	25/06/2025
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Accredited for compliance with ISO/IEC 17025 - Testing. Tests not covered by NATA are denoted with *	

Results Approved By

Amanda Chui, LC/Air Toxics Supervisor

Authorised By

Nancy Zhang, Laboratory Manager

PFAS in Water LOW LEVEL Short

Our Reference		384043-1	384043-2	384043-3
Your Reference	UNITS	Namoi Reservoir A25NA0100038	Namoi Reservoir Duplicate	Namoi Reservoir Field Blank
Barcode		A25NA0100038	-	-
Sample Site Code		N27	-	-
Date Sampled		23/06/2025	23/06/2025	23/06/2025
Type of sample		Water	Water	Water
Date prepared	-	25/06/2025	25/06/2025	25/06/2025
Date analysed	-	25/06/2025	25/06/2025	25/06/2025
Perfluorobutanesulfonic acid	µg/L	<0.001	<0.001	<0.001
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.004	0.004	<0.001
Perfluorooctanesulfonic acid PFOS	µg/L	0.006	0.006	<0.001
Perfluorooctanoic acid PFOA	µg/L	<0.001	<0.001	<0.001
6:2 FTS	µg/L	<0.001	<0.001	<0.001
8:2 FTS	µg/L	<0.002	<0.002	<0.002
Surrogate ¹³ C ₈ PFOS	%	81	84	82
Surrogate ¹³ C ₂ PFOA	%	107	110	111
Extracted ISTD ¹³ C ₃ PFBS	%	88	91	86
Extracted ISTD ¹⁸ O ₂ PFHxS	%	95	91	93
Extracted ISTD ¹³ C ₄ PFOS	%	95	94	95
Extracted ISTD ¹³ C ₄ PFOA	%	114	107	110
Extracted ISTD ¹³ C ₂ 6:2FTS	%	141	146	135
Extracted ISTD ¹³ C ₂ 8:2FTS	%	#	187	180
Total Positive PFHxS & PFOS	µg/L	0.0099	0.010	<0.001
Total Positive PFOA & PFOS	µg/L	0.006	0.006	<0.001
Total Positive PFAS	µg/L	0.01	0.010	<0.001

Method ID	Methodology Summary
Org-029	<p>Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.</p> <p>Analysis is undertaken with LC-MS/MS.</p> <p>PFAS results include the sum of branched and linear isomers where applicable.</p> <p>Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.</p> <p>Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.</p>

Client Reference: Narrabri Shire - Namoi Reservoir N27

QUALITY CONTROL: PFAS in Water LOW LEVEL Short						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	384043-2
Date prepared	-			25/06/2025	1	25/06/2025	25/06/2025		25/06/2025	25/06/2025
Date analysed	-			25/06/2025	1	25/06/2025	25/06/2025		25/06/2025	25/06/2025
Perfluorobutanesulfonic acid	µg/L	0.001	Org-029	<0.001	1	<0.001	<0.001	0	105	93
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.001	Org-029	<0.001	1	0.004	0.004	0	97	94
Perfluorooctanesulfonic acid PFOS	µg/L	0.001	Org-029	<0.001	1	0.006	0.006	0	101	95
Perfluorooctanoic acid PFOA	µg/L	0.001	Org-029	<0.001	1	<0.001	<0.001	0	92	102
6:2 FTS	µg/L	0.001	Org-029	<0.001	1	<0.001	<0.001	0	108	115
8:2 FTS	µg/L	0.002	Org-029	<0.002	1	<0.002	<0.002	0	101	97
Surrogate ¹³ C ₈ PFOS	%		Org-029	84	1	81	80	1	81	80
Surrogate ¹³ C ₂ PFOA	%		Org-029	106	1	107	103	4	110	109
Extracted ISTD ¹³ C ₃ PFBS	%		Org-029	94	1	88	88	0	82	93
Extracted ISTD ¹⁸ O ₂ PFHxS	%		Org-029	98	1	95	95	0	92	98
Extracted ISTD ¹³ C ₄ PFOS	%		Org-029	102	1	95	97	2	94	104
Extracted ISTD ¹³ C ₄ PFOA	%		Org-029	117	1	114	116	2	105	110
Extracted ISTD ¹³ C ₂ 6:2FTS	%		Org-029	142	1	141	143	1	128	136
Extracted ISTD ¹³ C ₂ 8:2FTS	%		Org-029	185	1	#	187		176	196

Result Definitions

NT	Not tested
NA	Test not required
INS	Insufficient sample for this test
PQL	Practical Quantitation Limit
<	Less than
>	Greater than
RPD	Relative Percent Difference
LCS	Laboratory Control Sample
NS	Not specified
NEPM	National Environmental Protection Measure
NR	Not Reported

Quality Control Definitions

Blank	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
Duplicate	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
Matrix Spike	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
LCS (Laboratory Control Sample)	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
Surrogate Spike	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.

Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Air volumes are typically provided by customers (often as flow rate(s) and sampling time(s) and/or simply volumes) sampled or exposure times (determines 'volume' passive badges are exposed to)). Hence in such circumstances the volume measurement is inevitably not covered by Envirolab's NATA accreditation. An exception may occur where Envirolab Newcastle does the sampling where accreditation exists for certain types of sampling and hence volume determination(s). Note air volumes are often used to determine concentrations for dust and/or analyses on filters, sorbents and in impingers. For canister sampling, the air volume is covered by Envirolab's NATA accreditation.

Urine Analysis - The BEI values listed are taken from the 2022 edition of "TLVs and BEIs Threshold Limits" by ACGIH.

Report Comments

For PFAS Extracted Internal Standards denoted with # or outside the 50-150% acceptance range, the respective target analyte results may be unaffected, in other circumstances the PQL has been raised to accommodate the outlier(s).

CERTIFICATE OF ANALYSIS 384044

Client Details

Client	NSW Health
Attention	Kwendy Cavanagh
Address	Locked Bag 2030, ST LEONARDS, NSW, 1590

Sample Details

Your Reference	<u>Narrabri Shire - Elizabeth Bore N30</u>
Number of Samples	3 Water
Date samples received	24/06/2025
Date completed instructions received	24/06/2025

Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

Please refer to the last page of this report for any comments relating to the results.

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Results Approved By

Amanda Chui, LC/Air Toxics Supervisor

Authorised By

Nancy Zhang, Laboratory Manager

PFAS in Water LOW LEVEL Short		
Our Reference		384044-1
Your Reference	UNITS	Elizabeth Bore A25NA0100036
Barcode		A25NA0100036
Sample Site Code		N30
Date Sampled		23/06/2025
Type of sample		Water
Date prepared	-	25/06/2025
Date analysed	-	25/06/2025
Perfluorobutanesulfonic acid	µg/L	<0.001
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.002
Perfluorooctanesulfonic acid PFOS	µg/L	0.005
Perfluorooctanoic acid PFOA	µg/L	<0.001
6:2 FTS	µg/L	<0.001
8:2 FTS	µg/L	<0.002
Surrogate ¹³ C ₈ PFOS	%	85
Surrogate ¹³ C ₂ PFOA	%	114
Extracted ISTD ¹³ C ₃ PFBS	%	88
Extracted ISTD ¹⁸ O ₂ PFHxS	%	87
Extracted ISTD ¹³ C ₄ PFOS	%	92
Extracted ISTD ¹³ C ₄ PFOA	%	114
Extracted ISTD ¹³ C ₂ 6:2FTS	%	161
Extracted ISTD ¹³ C ₂ 8:2FTS	%	177
Total Positive PFHxS & PFOS	µg/L	0.006
Total Positive PFOA & PFOS	µg/L	0.005
Total Positive PFAS	µg/L	0.006

Method ID	Methodology Summary
Org-029	<p>Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.</p> <p>Analysis is undertaken with LC-MS/MS.</p> <p>PFAS results include the sum of branched and linear isomers where applicable.</p> <p>Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.</p> <p>Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.</p>

QUALITY CONTROL: PFAS in Water LOW LEVEL Short						Duplicate		Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			25/06/2025	[NT]	[NT]	[NT]	[NT]	25/06/2025	[NT]
Date analysed	-			25/06/2025	[NT]	[NT]	[NT]	[NT]	25/06/2025	[NT]
Perfluorobutanesulfonic acid	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	100	[NT]
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	99	[NT]
Perfluorooctanesulfonic acid PFOS	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	99	[NT]
Perfluorooctanoic acid PFOA	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	90	[NT]
6:2 FTS	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	104	[NT]
8:2 FTS	µg/L	0.002	Org-029	<0.002	[NT]	[NT]	[NT]	[NT]	94	[NT]
Surrogate ¹³ C ₈ PFOS	%		Org-029	79	[NT]	[NT]	[NT]	[NT]	83	[NT]
Surrogate ¹³ C ₂ PFOA	%		Org-029	106	[NT]	[NT]	[NT]	[NT]	103	[NT]
Extracted ISTD ¹³ C ₃ PFBS	%		Org-029	101	[NT]	[NT]	[NT]	[NT]	90	[NT]
Extracted ISTD ¹⁸ O ₂ PFHxS	%		Org-029	96	[NT]	[NT]	[NT]	[NT]	90	[NT]
Extracted ISTD ¹³ C ₄ PFOS	%		Org-029	100	[NT]	[NT]	[NT]	[NT]	92	[NT]
Extracted ISTD ¹³ C ₄ PFOA	%		Org-029	124	[NT]	[NT]	[NT]	[NT]	115	[NT]
Extracted ISTD ¹³ C ₂ 6:2FTS	%		Org-029	155	[NT]	[NT]	[NT]	[NT]	134	[NT]
Extracted ISTD ¹³ C ₂ 8:2FTS	%		Org-029	191	[NT]	[NT]	[NT]	[NT]	177	[NT]

Result Definitions

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Air volumes are typically provided by customers (often as flow rate(s) and sampling time(s) and/or simply volumes) sampled or exposure times (determines 'volume' passive badges are exposed to)). Hence in such circumstances the volume measurement is inevitably not covered by Envirolab's NATA accreditation. An exception may occur where Envirolab Newcastle does the sampling where accreditation exists for certain types of sampling and hence volume determination(s). Note air volumes are often used to determine concentrations for dust and/or analyses on filters, sorbents and in impingers. For canister sampling, the air volume is covered by Envirolab's NATA accreditation.

Urine Analysis - The BEI values listed are taken from the 2022 edition of "TLVs and BEIs Threshold Limits" by ACGIH.

Report Comments

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CERTIFICATE OF ANALYSIS 384046

Client Details

Client	NSW Health
Attention	Kwendy Cavanagh
Address	Locked Bag 2030, ST LEONARDS, NSW, 1590

Sample Details

Your Reference	<u>Narrabri Shire - Tibberena Bore N28</u>
Number of Samples	3 Water
Date samples received	24/06/2025
Date completed instructions received	24/06/2025

Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

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Results Approved By

Amanda Chui, LC/Air Toxics Supervisor

Authorised By

Nancy Zhang, Laboratory Manager

PFAS in Water LOW LEVEL Short		
Our Reference		384046-1
Your Reference	UNITS	Tibberena Bore A25NA0100037
Barcode		A25NA0100037
Sample Site Code		N28
Date Sampled		23/06/2025
Type of sample		Water
Date prepared	-	25/06/2025
Date analysed	-	25/06/2025
Perfluorobutanesulfonic acid	µg/L	0.001
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.007
Perfluorooctanesulfonic acid PFOS	µg/L	0.011
Perfluorooctanoic acid PFOA	µg/L	<0.001
6:2 FTS	µg/L	<0.001
8:2 FTS	µg/L	<0.002
Surrogate ¹³ C ₈ PFOS	%	79
Surrogate ¹³ C ₂ PFOA	%	111
Extracted ISTD ¹³ C ₃ PFBS	%	99
Extracted ISTD ¹⁸ O ₂ PFHxS	%	97
Extracted ISTD ¹³ C ₄ PFOS	%	107
Extracted ISTD ¹³ C ₄ PFOA	%	123
Extracted ISTD ¹³ C ₂ 6:2FTS	%	166
Extracted ISTD ¹³ C ₂ 8:2FTS	%	#
Total Positive PFHxS & PFOS	µg/L	0.018
Total Positive PFOA & PFOS	µg/L	0.011
Total Positive PFAS	µg/L	0.019

Method ID	Methodology Summary
Org-029	<p>Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.</p> <p>Analysis is undertaken with LC-MS/MS.</p> <p>PFAS results include the sum of branched and linear isomers where applicable.</p> <p>Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.</p> <p>Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.</p>

Client Reference: Narrabri Shire - Tibberena Bore N28

QUALITY CONTROL: PFAS in Water LOW LEVEL Short					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			25/06/2025	[NT]	[NT]	[NT]	[NT]	25/06/2025	[NT]
Date analysed	-			25/06/2025	[NT]	[NT]	[NT]	[NT]	25/06/2025	[NT]
Perfluorobutanesulfonic acid	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	100	[NT]
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	99	[NT]
Perfluorooctanesulfonic acid PFOS	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	99	[NT]
Perfluorooctanoic acid PFOA	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	90	[NT]
6:2 FTS	µg/L	0.001	Org-029	<0.001	[NT]	[NT]	[NT]	[NT]	104	[NT]
8:2 FTS	µg/L	0.002	Org-029	<0.002	[NT]	[NT]	[NT]	[NT]	94	[NT]
Surrogate ¹³ C ₈ PFOS	%		Org-029	79	[NT]	[NT]	[NT]	[NT]	83	[NT]
Surrogate ¹³ C ₂ PFOA	%		Org-029	106	[NT]	[NT]	[NT]	[NT]	103	[NT]
Extracted ISTD ¹³ C ₃ PFBS	%		Org-029	101	[NT]	[NT]	[NT]	[NT]	90	[NT]
Extracted ISTD ¹⁸ O ₂ PFHxS	%		Org-029	96	[NT]	[NT]	[NT]	[NT]	90	[NT]
Extracted ISTD ¹³ C ₄ PFOS	%		Org-029	100	[NT]	[NT]	[NT]	[NT]	92	[NT]
Extracted ISTD ¹³ C ₄ PFOA	%		Org-029	124	[NT]	[NT]	[NT]	[NT]	115	[NT]
Extracted ISTD ¹³ C ₂ 6:2FTS	%		Org-029	155	[NT]	[NT]	[NT]	[NT]	134	[NT]
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Result Definitions

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Surrogate Spike	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.

Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Air volumes are typically provided by customers (often as flow rate(s) and sampling time(s) and/or simply volumes) sampled or exposure times (determines 'volume' passive badges are exposed to)). Hence in such circumstances the volume measurement is inevitably not covered by Envirolab's NATA accreditation. An exception may occur where Envirolab Newcastle does the sampling where accreditation exists for certain types of sampling and hence volume determination(s). Note air volumes are often used to determine concentrations for dust and/or analyses on filters, sorbents and in impingers. For canister sampling, the air volume is covered by Envirolab's NATA accreditation.

Urine Analysis - The BEI values listed are taken from the 2022 edition of "TLVs and BEIs Threshold Limits" by ACGIH.

Report Comments

For PFAS Extracted Internal Standards denoted with # or outside the 50-150% acceptance range, the respective target analyte results may be unaffected, in other circumstances the PQL has been raised to accommodate the outlier(s).